

# Gut and Root Microbiota Commonalities

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**Animal guts and plant roots have absorption roles for nutrient uptake and converge in harboring large, complex, and dynamic groups of microbes that participate in degradation or modification of nutrients and other substances. Gut and root bacteria regulate host gene expression, provide metabolic capabilities, essential nutrients, and protection against pathogens, and seem to share evolutionary trends.**

Guts and roots are inhabited by many different bacteria (1–5), archaea (6–12), and viruses (13–16), as well as by eukaryotes (17–20), with some of them containing bacteria of their own (21–24). Variations in gut microbiota respond to age (25–28), diet (29–31), or species (32). Most insects have dozens of microbial species in their guts, while mammalian guts may contain thousands. Herbivores exhibit the largest diversity (32, 33), including probably plant-associated bacteria, especially endophytes (34) that, by being inside plant tissues, may survive stomach digestion. Transiting diet-borne bacteria may contribute to gut metabolic capacities. Different soil types, moisture (35), plant genotypes (36), age (37), and root lysates, secretions, or exudates (38) are determinants of root microbiotas. Factors that determine root exudates, such as availability of inorganic nutrients, temperature, light intensity, O<sub>2</sub>/CO<sub>2</sub> level, or root damage, may indirectly affect root microbiotas (39). The presence of pathogens induces changes in microbiota composition in roots and guts (40, 41).

Guts and roots have large surface areas, with microvilli and folds or root hairs in some parts. Both roots and guts are structured, nonhomogenous habitats with pH, nutrient, water, and oxygen differential levels or gradients. Gradients would favor colonization by distinct bacteria that are more successful in some root or gut regions. In consequence, the multiple microhabitats that exist in roots and guts contribute to high species richness (42, 43). Different conditions are found in the cecum and distal colon in humans, with cecal and colon microbiotas containing a larger proportion of facultative anaerobes (44). Colon mucosal folds exhibit particular bacteria adapted to colonic conditions and maybe to mucin degradation (45). Some insects have specialized structures in their gut, such as midgut sacs and tubular outgrowths called ceca or crypts, in which they harbor specific bacteria (46), and others with less-complex guts also have pH and oxygen gradients in their guts (47). A steep oxygen gradient including an anaerobic root environment in water-saturated roots parallels the gut oxygen gradient and anaerobic gut systems. Clostridia, and especially members of the family *Ruminococcaceae*, are more prevalent than other anaerobes and methanogens, a trend which is similar in the different gut systems (48). These communities take care of the degradation of the complex organic matter in the outer root layers. Some gut and root acid-tolerant bacteria can modify their environment by lowering the pH when producing diverse acids (49, 50). Along the roots, there are physiological differences, and their exudates are secreted differentially at the apical meristem, root cap, or root hairs (42), creating different microhabitats. A single *Burkholderia* strain colonizes only discrete root regions

(51), and different burkholderias were found at different soil depths (37).

“*Arabidopsis thaliana* root microbiome might assemble by core ecological principles similar to those shaping the mammalian microbiome in which core phylum level enterotypes provide broad metabolic potential combined with modest levels of host genotype-dependent associations” (35). Metacommunity theory may be applied to root microbiotas, as has been used to explain the assembly of the gut microbial community (52). Metacommunity theory is based on the concept of discontinuous patches and interactions that can satisfactorily describe bacterial patchy colonization of roots. Future applications of these concepts will assert their usefulness.

Remarkably, there are individual-to-individual variations in bacterial composition of the gut (2, 53) and roots (54). Individual differences may be due to genetic differences and stochastic colonization processes (52). Limited patterns (enterotypes) in relation to stratified variation were distinguished in human and insect gut microbiotas (2, 55); however, it is controversial if there are only a few enterotypes in humans or gradients of diversity (28). In plants, similar bacterial genera are recurrently isolated from rhizospheres (soil surrounding roots affected by plants) or roots (34, 56). In roots, *Rhizobium* strain diversity with functional differentiation is high (57). Strain variability in vitamin production has been detected among gut bifidobacteria (reviewed in reference 58). Similarly, lactobacilli (reviewed in reference 59) are a heterogeneous group of bacteria with partly probiotic character which have considerable variation in terms of molecular characteristics and preferred natural habitats.

With few exceptions (see below), the gut microbiota is different from that of other host organs, and similarly, the root microbiota shares only some bacteria with those of other plant organs.

## ENVIRONMENTAL AND MATERNAL ACQUISITION

Root and gut microorganisms are usually acquired from the environment. Roots are colonized by bulk soil microorganisms at-

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tracted by chemotaxis and enriched by nutrients secreted by the roots in the rhizosphere. Animals also acquire their gut microbiota from their environment after they are born (60). In a few cases, microorganisms can be transferred vertically from mother to progenies. Endophytes present in plant seeds may subsequently colonize the roots and the rhizosphere. *Enterobacter asburiae*, found in maize kernels, is able to exit the roots and colonize the rhizosphere after the plant has established (61). Other seed bacteria do the same (54, 62). Animals can also acquire their gut microbiota from their mothers after being born, but there are cases of paternal transmission of symbionts, as in malaria vectors (63). Maternal transmission may occur before birth (64–66). When mammals are breast-fed, they acquire microorganisms that are present in the milk or on the mother's skin (67–69). Some stink-bug larvae acquire their mother's gut bacteria from contaminated eggs, by coprophagy, or by capsule-mediated transmission just after they have hatched (46). In view of the vertical and environmental transmission of root and gut microbes, gnotobiotic animals or plants are needed to clearly evaluate the effects of selected strains on hosts.

#### FUNCTIONAL REDUNDANCY AND ROLE OF MINORITIES

It seems that different microbiota composition may lead to the same and stable function. This may apply to gut and root bacteria and has been found to be true in methanogenic reactors (70). Similar degrading capacities are found in different gut bacteria (reviewed in reference 71). In roots, many different bacterial genera and species produce hormones, auxins, cytokinins, or gibberellins (reviewed in references 56 and 72). Our research group found that riboflavin is produced and excreted by different strains from several species of *Methylobacterium*, *Rhizobium*, *Sinorhizobium*, and *Bacillus*, both in rice and alfalfa root exudates and in pure cultures in minimal medium (our unpublished data). *In vitro* excretion of riboflavin by a large diversity of bacteria, including *Chromobacterium violaceum* and *Pantoea agglomerans*, was reported earlier (73), and both riboflavin and lumichrome (which is derived from riboflavin) stimulate root respiration (74). Additionally, many different plant-associated bacteria inhibit pathogenic fungi or bacteria (reviewed in reference 56).

Minority species present in the microbiota may help cover some of the host-specific needs. Methanogens, methylotrophs, and nitrogen-fixing bacteria are minor components in guts and rhizospheres (11, 75–78); however, they have important ecological roles. In some roots and guts, nitrogen fixation provides nitrogen to plants (79) and insects (80–82).

#### GUT AND ROOT BACTERIA ENHANCE THE METABOLIC CAPACITIES OF THEIR HOSTS

It is remarkable that gut bacteria are rich in sugar hydrolases (83) and other catabolic genes, such as those for tannin (84), cholesterol (85), or mucin (gut glycosylated proteins) (86). Similarly, capacities to degrade polyphenols, polysaccharides, protocatechuate, and proteins and to solubilize phosphate and weather rocks (50, 54, 87, 88) are prevalent among different rhizospheric bacteria. Mimosine-degrading bacteria are found in mimosa plants that produce mimosine (89), and cows that have such bacteria in their rumen are capable of degrading it (90). Alginate-degrading bacteria are found in abalone and human guts of algae consumers in Japan (91). The outstanding degrading capacities of root bacteria are the basis of rhizoremediation of polluting substances (92, 93)

and are also evidenced in medical drug transformation or degradation in the human gut (94–96). Interestingly, in bioremediation, the abilities of bacteria to degrade soil pollutants may be triggered by flavonoids (97).

Gut and rhizospheric bacteria produce vitamins as riboflavin, as stated above. Vitamin B<sub>12</sub> is an exclusive product of prokaryotes (98), and it is produced by plant root and gut bacteria (99–102). Essential amino acids and vitamins B and K are produced by gut bacteria (reviewed in reference 58). An alcohol dehydrogenase from the commensal bacterium *Acetobacter pomorum* modulates *Drosophila* developmental and metabolic homeostasis via insulin signaling (103). While root bacteria produce plant hormones that have effects on plant growth (reviewed in reference 56), gut bacteria seem to regulate animal behavior (104, 105).

#### GUT AND ROOT MICROBIOTAS COMPETE WITH PATHOGENS

Gut and root microbiotas suppress pathogens (reviewed in references 56 and 106). The human control of root bacteria has been envisaged as a manner to promote plant growth and health with benefits to agriculture (93, 107). Bacterial inoculants in agriculture and forestry are considered equivalent to probiotics (beneficial microbes provided as supplements) for animal health. Probiotics stimulate host defense systems and the competitive exclusion of pathogens, as plant growth-promoting rhizobacteria do (108). Seeds may harbor a reservoir of probiotics for their seedlings (54, 109). Prebiotics are added nutrients used to stimulate desirable bacteria in humans (110). We may even speculate that prebiotics were invented by roots, as some substances from their exudates stimulate bacterial growth selectively (89, 111, 112).

For over one hundred years, inoculants have been provided to plants in agricultural fields with variable success. Recently, a large number of commercial products whose effects are not always desirable have appeared to promote plant growth. Similarly, an increased number of probiotics and prebiotics whose effects have not been completely evaluated in different human populations are coming to the market. Gut gene expression in response to probiotics varies from person to person (113). In many cases, clinical benefits have been obtained in patients with specific probiotic strains (114).

Experience with plants has shown that appropriate use and regulation of probiotics (inoculants) is difficult to achieve. Undesirable genetic characteristics, such as denitrifying capacities, have been identified among inoculants (115). Strains used as probiotics should not contain glucosaminidase or glucuronidase genes that seem to have roles in producing toxic substances in the gut (reviewed in reference 116), but these recommendations may not be easily followed.

#### SIMILAR BACTERIUM-HOST INTERACTIONS IN GUTS AND ROOTS

**Differential gene expression of bacteria in hosts.** Bacterium-plant interactions have been studied for many years, and a molecular ping pong between rhizobia and plants that may serve as a model to analyze insect or human gut symbioses is known (reviewed in references 1 and 117). In rhizobium-plant molecular dialogue, *Rhizobium* NodD receptors, which bind root exudate molecules, function as transcriptional regulators that induce the expression of several genes, including *nod* genes and secretion systems (reviewed in references 117 and 118). Extrusion pumps are inducible by flavonoids that are present in root exudates but

do not require NodD genes (119). Many ABC transporter systems are induced by the respective substrate or other molecules from roots (111, 120).

In roots, bacteria have a differential gene expression that supposedly allows them to adapt to the root environment. Genes involved in root exudate usage, root attachment, and survival are induced in bacteria colonizing roots (120, 121). *In vitro* expression technology (IVET) (122), proteomic analysis, microarray and RNA Seq transcriptomics, and genetic analysis have revealed rhizobial (120, 121, 123), *Pseudomonas* (124, 125), *Streptomyces* (126), and other bacterial genes expressed on roots or rhizospheres. Similarly, bacteria may differentially express genes when in guts. Gut bacteria are exposed to bile salts that solubilize diet fat, have antimicrobial activities (127), and regulate bacterial gene expression. An efflux transporter of the multidrug resistance type (MDR) was induced in *Bifidobacterium* by bile (128). Different bile substances have been identified to control gene expression in bifidobacteria (129). Other bile-inducible genes have been found in *Lactobacillus plantarum* (130). Lastly, human gut bacteria transform bile salts (131). Gut bacteria can also modify dietary flavonoids (132) that have significant effects on animal physiology. Analogously, in roots, flavonoids produced by plants are signal molecules in bacteria (133) and are also transformed by bacteria *in vitro*, though this has not been shown *in vivo*. Plant phytoalexins are antimicrobials that are expelled from *Rhizobium etli*, *Bradyrhizobium japonicum*, and *Agrobacterium* by MDR efflux pumps that are inducible by root-exuded flavonoids (20, 119, 134).

Interestingly, gut and root microbiotas may follow the circadian cycles of their hosts. This was observed in nitrogen-fixing bacteria that fixed more during the daytime on rice roots (135). Epithelial cell proliferation, gastrointestinal motility, and other gut processes follow biological rhythms. In the gastrointestinal tract, there are large amounts of melatonin, which is a key hormone in the clock biological regulation (136). The Burmese python's microbiota is responsive to host cycles of feeding and fasting (137).

**Host gene expression regulated by microbiotas.** Outstandingly, gut and root bacteria modify gene expression in animal (138, 139) and plant (140) hosts, respectively. Gut gene expression is also modified by probiotics (113) that modify gut bacterial gene expression as well (141). Gut genes expressed in the presence of the gut bacterium *Bacteroides thetaiotaomicron* are involved in xenobiotic catabolism, in angiogenesis, in gut barrier epithelium maintenance, and in immunity development (139), with very complex host molecular responses (142).

Plants and humans can sense bacterially produced acylhomoserine lactones (AHLs), different volatiles, microbe-associated molecular patterns (MAMPS) (72, 143), and other bacterial molecules unknown at present. Root gene expression is differently modified by acylhomoserine lactones from pathogenic or symbiotic bacteria (144). In turn, plant products may act like quorum-sensing signals in bacteria (145). In recent years, specific regulatory roles of *N*-acylhomoserine lactones have become apparent, because plants responded with either a systemic resistance response or a hormonal regulated growth response to the presence of AHL-producing bacteria colonizing the root surface. Also in the animal/human systems, a specific perception of AHL compounds, produced by Gram-negative, mostly pathogenic bacteria, was found in many tissues, including the gut system, leading to immu-

nomodulatory effects (146). In plants, root genes induced by rhizospheric bacteria are involved in oxidative and defense responses, in plant secondary metabolism, or in signaling (140). Plants may detect bacterial cyclopeptides through auxin sensing pathways (147). In a more specialized symbiosis, a cascade of signaling processes occurs inside root cells in the presence of rhizobia or Nod factors (148).

**Control of microbiotas.** A *Drosophila* mutant with increased levels of antimicrobial peptides showed deregulated balances of gut populations (149), with smaller numbers of *Commensalibacter intestini* (an acetic acid bacterium present in normal gut) bacteria (150) and increased numbers of *Gluconobacter morbifer* cells that caused gut cell apoptosis and early insect death (149). It is interesting to note that *C. intestini* antagonizes *G. morbifer*, which is a normal gut member, but with detrimental effects when present in large numbers; thus, *C. intestini* contributes to gut homeostasis and host fitness (151). Similarly, among root microbiotas, there are plant-pathogenic bacteria that normally would not affect the plants when kept in low numbers by other plant community strains or plant antimicrobials. Lipopolysaccharide *Rhizobium* mutants that were more sensitive to maize antimicrobial benzoxazinones had reduced rhizospheric colonization (152). Antimicrobial peptides constitute a line of defense in plants as effectors of innate immunity and regulate not only bacteria but also methanogenic archaea in guts (153). Gut immunity determines bacterial composition; reciprocally, bacteria modulate host immunity in guts (154, 155). Carbohydrate binding proteins (lectins) from guts and roots bind bacteria, form aggregates, and may have antibacterial effects (156, 157).

In addition to bacterium-host interactions, bacterium-bacterium interactions may determine community composition and its function (158). Those that occur in the mouth (159) may guide research in gut and root symbioses. In *Rhizobium*, mutants in quorum sensing are affected in rhizosphere colonization (160). Acylhomoserine lactones may be degraded by rhizospheric bacteria causing interference with quorum signals that regulate gene expression in other bacteria (161). This may have a role in protecting plants from pathogens but may also affect mutualistic interactions.

## EVOLUTIONARY PATHWAYS

**Lateral gene transfer in guts and roots.** In roots, root nodules, and guts, lateral transfer of genetic material between different bacteria has been evidenced (2, 162, 163), seemingly promoted by close contacts in high-density populations. The presence of similar catabolic or antibiotic resistance genes in various gut bacterial genera has been explained as acquisitions by lateral gene transfers (91). It has been suggested that starch catabolism genes have been transferred from gut to bacteria (164).

There are many more phages than bacteria in the gut (13), and some may be involved in lateral gene transfer among gut bacteria (165). Lateral transfer of genetic material is mediated by plasmids or genomic island mobilization in rhizobia and other rhizospheric bacteria (54, 166), but phages may have a role as well.

**Specialized symbiont evolution from root and gut bacteria.** It has been suggested that gut bacteria gave rise to endosymbiotic bacteria in insects (167) based on similarities of gut bacteria and insect endosymbionts (168). Correspondingly, rhizospheric bacteria may have preceded nodule and endophytic bacteria in plants (169). Insect endosymbionts and nodule rhizobia are selected



symbionts that occupy intracellularly host-specialized structures and attain high numbers with a determined functional role. However, transmission modes of plant- and insect-specialized symbionts (reviewed in reference 46) and their genome sizes (rhizobial genome sizes reviewed in references 121 and 170) are different.

## CONCLUSIONS

The comparison of plant and gut microbial ecologies may help to guide research toward the understanding of such complex symbioses. Literature on the subject is so extensive that only a few references were used to illustrate the commonalities of gut and root microbiotas. Interested readers are referred to recent literature (171–175). Plants use their “guts” (roots) outwards, and this simplifies their study in comparison to study of animal guts. Gut and root microbiotas significantly impact health, development, and fitness of their respective hosts.

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## REFERENCES

- Badri DV, Weir TL, van der Lelie D, Vivanco JM. 2009. Rhizosphere chemical dialogues: plant-microbe interactions. *Curr. Opin. Biotechnol.* 20:642–650.
- Dillon RJ, Dillon VM. 2004. The gut bacteria of insects: nonpathogenic interactions. *Annu. Rev. Entomol.* 49:71–92.
- Kurokawa K, Itoh T, Kuwahara T, Oshima K, Toh H, Toyoda A, Takami H, Morita H, Sharma VK, Srivastava TP, Taylor TD, Noguchi H, Mori H, Ogura Y, Ehrlich DS, Itoh K, Takagi T, Sakaki Y, Hayashi T, Hattori M. 2007. Comparative metagenomics revealed commonly enriched gene sets in human gut microbiomes. *DNA Res.* 14:169–181.
- Marchesi JR. 2010. Prokaryotic and eukaryotic diversity of the human gut. *Adv. Appl. Microbiol.* 72:43–62.
- Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JI. 2007. The human microbiome project. *Nature* 449:804–810.
- Donovan SE, Purdy KJ, Kane MD, Eggleton P. 2004. Comparison of *Euryarchaea* strains in the guts and food-soil of the soil-feeding termite *Cubitermes fungifaber* across different soil types. *Appl. Environ. Microbiol.* 70:3884–3892.
- Fricke WF, Seedorf H, Henne A, Krüer M, Liesegang H, Hedderich R, Gottschalk G, Thauer RK. 2006. The genome sequence of *Methanospirillum hutchinsonii* reveals why this human intestinal archaeon is restricted to methanol and H<sub>2</sub> for methane formation and ATP synthesis. *J. Bacteriol.* 188:642–658.
- Friedrich MW, Scmitt-Wagner D, Leuders T, Brune A. 2001. Axial differences in community structure of *Crenarchaeota* and *Euryarchaeota* in the highly compartmentalized gut of the soil-feeding termite *Cubitermes orthognathus*. *Appl. Environ. Microbiol.* 67:4880–4890.
- Hara K, Shinzato N, Seo M, Oshima T, Yamagishi A. 2002. Phylogenetic analysis of symbiotic archaea living in the gut of xylophagous cockroaches. *Microbes Environ.* 17:185–190.
- Horz HP, Conrads G. 2010. The discussion goes on: what is the role of *Euryarchaeota* in humans? *Archaea* 2010:967271.
- Jarrell KE, Walters AD, Bochiwal C, Borgia JM, Dickinson T, Chong JP. 2011. Major players on the microbial stage: why archaea are important. *Microbiology* 157:919–936.
- Simon HM, Dodsworth JA, Goodman RM. 2000. *Crenarchaeota* colonize terrestrial plant roots. *Environ. Microbiol.* 2:495–505.
- Minot S, Grunberg S, Wu GD, Lewis JD, Bushman FD. 2012. Hyper-variable loci in the human gut virome. *Proc. Natl. Acad. Sci. U. S. A.* 109:3962–3966.
- Minot S, Sinha R, Chen J, Li H, Keilbaugh SA, Wu GD, Lewis JD, Bushman FD. 2011. The human gut virome: inter-individual variation and dynamic response to diet. *Genome Res.* 21:1616–1625.
- Reyes A, Haynes M, Hanson N, Angly FE, Heath AC, Rohwer F, Gordon JI. 2010. Viruses in the faecal microbiota of monozygotic twins and their mothers. *Nature* 466:334–338.
- Swanson MM, Fraser G, Daniell TJ, Torrance L, Gregory PJ, Taliansky M. 2009. Viruses in soils: morphological diversity and abundance in the rhizosphere. *Ann. Appl. Biol.* 155:51–60.
- Nam YD, Chang HW, Kim KH, Roh SW, Kim MS, Jung MJ, Lee SW, Kim JY, Yoon JH, Bae JW. 2008. Bacterial, archaeal, and eukaryal diversity in the intestines of Korean people. *J. Microbiol.* 46:491–501.
- Pandey PK, Siddharth J, Verma P, Bavdekar A, Patole MS, Shouche YS. 2012. Molecular typing of fecal eukaryotic microbiota of human infants and their respective mothers. *J. Biosci.* 37:221–226.
- Parfrey LW, Walters WA, Knight R. 2011. Microbial eukaryotes in the human microbiome: ecology, evolution, and future directions. *Front. Microbiol.* 2:153.
- Parniske M. 2008. Arbuscular mycorrhiza: the mother of plant root endosymbioses. *Nat. Rev. Microbiol.* 6:763–775.
- Bertaux J, Schmid M, Chemidlin Prevost-Boure N, Churin JL, Hartmann A, Garbaye J, Frey-Klett P. 2003. *In situ* identification of intracellular bacteria related to *Paenibacillus* spp. in the mycelium of the ectomycorrhizal fungus *Laccaria bicolor* S238N. *Appl. Environ. Microbiol.* 69:4243–4248.
- Bianciotto V, Lumini E, Bonfante P, Vandamme P. 2003. ‘*Candidatus Glomeribacter gigasporarum*’ gen. nov., sp. nov., an endosymbiont of arbuscular mycorrhizal fungi. *Int. J. Syst. Evol. Microbiol.* 53:121–124.
- Scheublin TR, Sanders IR, Keel C, van der Meer JR. 2010. Characterisation of microbial communities colonising the hyphal surfaces of arbuscular mycorrhizal fungi. *ISME J.* 4:752–763.
- Stingl U, Radek R, Yang H, Brune A. 2005. “Endomicrobia”: cytoplasmic symbionts of termite gut protozoa form a separate phylum of prokaryotes. *Appl. Environ. Microbiol.* 71:1473–1479.
- Biagi E, Candela M, Fairweather-Tait S, Franceschi C, Brigidi P. 2012. Ageing of the human metaorganism: the microbial counterpart. *Age* 34:247–267.
- Mihajlovski A, Doré J, Levenez F, Alric M, Brugère JF. 2010. Molecular evaluation of the human gut methanogenic archaeal microbiota reveals an age-associated increase of the diversity. *Environ. Microbiol. Rep.* 2:272–280.
- Tiihonen K, Ouwehand AC, Rautonen N. 2010. Human intestinal microbiota and healthy ageing. *Ageing Res. Rev.* 9:107–116.
- Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, Hidalgo G, Baldassano RN, Anokhin AP, Heath AC, Warner B, Reeder J, Kuczynski J, Caporaso JG, Lozupone CA, Lauber C, Clemente JC, Knights D, Knight R, Gordon JI. 2012. Human gut microbiome viewed across age and geography. *Nature* 486:222–227.
- Muegge BD, Kuczynski J, Knights D, Clemente JC, González A, Fontana L, Henrissat B, Knight R, Gordon JI. 2011. Diet drives convergence in gut microbiome functions across mammalian phylogeny and within humans. *Science* 332:970–974.
- Turnbaugh PJ, Ridaura VK, Faith JJ, Rey FE, Knight R, Gordon JI. 2009. The effect of diet on the human gut microbiome: a metagenomic analysis in humanized gnotobiotic mice. *Sci. Transl. Med.* 1:6ra14. doi: 10.1126/scitranslmed.3000322.
- Hooper LV, Midtvedt T, Gordon JI. 2002. How host-microbial interactions shape the nutrient environment of the mammalian intestine. *Annu. Rev. Nutr.* 22:283–307.
- Ley RE, Hamady M, Lozupone C, Turnbaugh PJ, Ramey RR, Bircher JS, Schlegel ML, Tucker TA, Schrenzel MD, Knight R, Gordon JI. 2008. Evolution of mammals and their gut microbes. *Science* 320:1647–1651.
- Hong PY, Wheeler E, Cann IKO, Mackie RI. 2011. Phylogenetic analysis of the fecal microbial community in herbivorous land and marine iguanas of the Galápagos Islands using 16S rRNA-based pyrosequencing. *ISME J.* 5:1461–1470.
- Rosenblueth M, Martínez-Romero E. 2006. Bacterial endophytes and their interactions with hosts. *Mol. Plant Microbe Interact.* 19:827–837.
- Lundberg DS, Lebeis SL, Paredes SH, Yourstone S, Gehring J, Malfatti S, Tremblay J, Engelbrektson A, Kunin V, del Rio TG, Edgar RC, Eickhorst T, Ley RE, Hugenholtz P, Tringe SG, Dangl JL. 2012. Defining the core *Arabidopsis thaliana* root microbiome. *Nature* 488:86–90.
- Hartmann A, Schmid M, van Tuinen D, Berg G. 2009. Plant-driven selection of microbes. *Plant Soil* 321:235–257.
- Chiarini L, Giovannelli V, Bevivino A, Dalmastrì C, Tabacchioni S. 2000. Different portions of the maize root system host *Burkholderia ce-*

- pacia* populations with different degrees of genetic polymorphism. *Environ. Microbiol.* 2:111–118.
38. Doornbos RF, van Loon LC, Bakker PAHM. 2012. Impact of root exudates and plant defense signaling on bacterial communities in the rhizosphere. A review. *Agron. Sustain. Dev.* 32:227–243.
  39. Rovira AD. 1969. Plant root exudates. *Bot. Rev.* 35:35–57.
  40. Chow J, Lee SM, Shen Y, Khosravi A, Mazmanian SK. 2010. Host-bacterial symbiosis in health and disease. *Adv. Immunol.* 107:243–274.
  41. Raaijmakers JM, Paulitz TC, Steinberg C, Alabouvette C, Moenne-Loccoz Y. 2009. The rhizosphere: a playground and battlefield for soil-borne pathogens and beneficial microorganisms. *Plant Soil* 321:341–361.
  42. Bertin C, Yang X, Weston LA. 2003. The role of root exudates and allelochemicals in the rhizosphere. *Plant Soil* 256:67–83.
  43. Turrone F, Marchesi JR, Foroni E, Gueimonde M, Shanahan F, Margolles A, van Sinderen D, Ventura M. 2009. Microbiomic analysis of the bifidobacterial population in the human distal gut free. *ISME J.* 3:745–751.
  44. Marteau P, Pochart P, Doré J, Béra-Maillet C, Bernalier A, Corthier G. 2001. Comparative study of bacterial groups within the human cecal and fecal microbiota. *Appl. Environ. Microbiol.* 67:4939–4942.
  45. Nava GM, Friedrichsen HJ, Stappenbeck TS. 2011. Spatial organization of intestinal microbiota in the mouse ascending colon. *ISME J.* 5:627–638.
  46. Kikuchi Y, Hosokawa T, Fukatsu T. 2008. Diversity of bacterial symbiosis in stinkbugs, p 39–63. *In* Dijk TV (ed), *Microbial Ecology Research Trends*. Nova Science Publishers Inc., New York, NY.
  47. Brune A, Emerson D, Breznak JA. 1995. The termite gut microflora as an oxygen sink—microelectrode determination of oxygen and pH gradients in guts of lower and higher termites. *Appl. Environ. Microbiol.* 61:2681–2687.
  48. Timmers RA, Rothballer M, Strik DP, Engel M, Schulz S, Schloter M, Hartmann A, Hamelers B, Buisman C. 2012. Microbial community structure elucidates performance of *Glyceria maxima* plant microbial fuel cell. *Appl. Microbiol. Biotechnol.* 94:537–548.
  49. Asahara T, Shimizu K, Nomoto K, Hamabata T, Ozawa A, Takeda Y. 2004. Probiotic bifidobacteria protect mice from lethal infection with Shiga toxin-producing *Escherichia coli* O157:H7. *Infect. Immun.* 72:2240–2247.
  50. Rodríguez H, Gonzalez T, Goire I, Bashan Y. 2004. Gluconic acid production and phosphate solubilization by the plant growth-promoting bacterium *Azospirillum* spp. *Naturwissenschaften* 91:552–555.
  51. Sharma S, Sharma S, Singh RK, Vaishampayan A. 2008. Colonization behavior of bacterium *Burkholderia cepacia* inside the *Oryza sativa* roots visualized using green fluorescent protein reporter. *World J. Microbiol. Biotechnol.* 24:1169–1175.
  52. Costello EK, Stagaman K, Dethlefsen L, Bohannan BJM, Relman DA. 2012. The application of ecological theory toward an understanding of the human microbiome. *Science* 336:1255–1262.
  53. Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, Gill SR, Nelson KE, Relman DA. 2005. Diversity of the human intestinal microbial flora. *Science* 308:1635–1638.
  54. López-López A, Rogel MA, Ormeño-Orrillo E, Martínez-Romero J, Martínez-Romero E. 2010. *Phaseolus vulgaris* seed-borne endophytic community with novel bacterial species such as *Rhizobium endophyticum* sp. nov. *Syst. Appl. Microbiol.* 33:322–327.
  55. Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, Fernandes GR, Tap J, Bruls T, Batto JM, Bertalan M, Borruel N, Casellas F, Fernandez L, Gautier L, Hansen T, Hattori M, Hayashi T, Kleerebezem M, Kurokawa K, Leclerc M, Levenez F, Manichanh C, Nielsen HB, Nielsen T, Pons N, Poulain J, Qin J, Sicheritz-Ponten T, Tims S, Torrents D, Ugarte E, Zoetendal EG, Wang J, Guarner F, Pedersen O, de Vos WM, Brunak S, Doré J, Weissenbach J, Ehrlich SD, Bork P. 2011. Enterotypes of the human gut microbiome. *Nature* 473:174–180.
  56. Friesen ML, Porter SS, Stark SC, von Wettberg EJ, Sachs JL, Martínez-Romero E. 2011. Microbially mediated plant functional traits. *Annu. Rev. Ecol. Evol. Syst.* 42:23–46.
  57. Rosenblueth M, Martínez-Romero E. 2004. *Rhizobium etli* endophytic populations and their competitiveness for root maize colonization. *Arch. Microbiol.* 181:337–344.
  58. Macfarlane S, Macfarlane GT. 2003. Food and the large intestine, p 24–51. *In* Fuller R, Perdigon G (ed), *Gut flora, nutrition, immunity and health*. Blackwell Publishing, Oxford, United Kingdom.
  59. Kleerebezem M, Hols P, Bernard E, Rolain T, Zhou M, Siezen RJ, Bron PA. 2010. The extracellular biology of the lactobacilli. *FEMS Microbiol. Rev.* 34:199–230.
  60. Kikuchi Y, Hosokawa T, Fukatsu T. 2011. An ancient but promiscuous host-symbiont association between *Burkholderia* gut symbionts and their heteropteran hosts. *ISME J.* 5:446–460.
  61. Johnston-Monje D, Raizada MN. 2011. Conservation and diversity of seed associated endophytes in *Zea* across boundaries of evolution, ethnography and ecology. *PLoS One* 6:e20396. doi:10.1371/journal.pone.0020396.
  62. Pereira P, Ibáñez F, Rosenblueth M, Etcheverry M, Martínez-Romero E. 2011. Analysis of the bacterial diversity associated with the roots of maize (*Zea mays* L.) through culture-dependent and culture-independent methods. *ISRN Ecol.* 2011:938546. doi:10.5402/2011/938546.
  63. Damiani C, Ricci I, Crotti E, Rossi P, Rizzi A, Scuppa P, Esposito F, Bandi C, Daffonchio D, Favia G. 2008. Paternal transmission of symbiotic bacteria in malaria vectors. *Curr. Biol.* 18:R1087–R1088.
  64. Jiménez E, Fernández L, Marín ML, Martín R, Odriozola JM, Nueno-Palop C, Narbad A, Olivares M, Xaus J, Rodríguez JM. 2005. Isolation of commensal bacteria from umbilical cord blood of healthy neonates born by cesarean section. *Curr. Microbiol.* 51:270–274.
  65. Jiménez E, Marín ML, Martín R, Odriozola JM, Olivares M, Xaus J, Fernández L, Rodríguez JM. 2008. Is meconium from healthy newborns actually sterile? *Res. Microbiol.* 159:187–193.
  66. Mshvildadze M, Neu J, Shuster J, Theriaque D, Li N, Mai V. 2010. Intestinal microbial ecology in premature infants assessed with non-culture-based techniques. *J. Pediatr.* 156:20–25.
  67. Hunt KM, Foster JA, Forney LJ, Schütte UM, Beck DL, Abdo Z, Fox LK, Williams JE, McGuire MK, McGuire MA. 2011. Characterization of the diversity and temporal stability of bacterial communities in human milk. *PLoS One* 6:e21313. doi:10.1371/journal.pone.0021313.
  68. Martín R, Jiménez E, Heilig H, Fernández L, Marín ML, Zoetendal EG, Rodríguez JM. 2009. Isolation of bifidobacteria from breast milk and assessment of the bifidobacterial population by PCR-denaturing gradient gel electrophoresis and quantitative real-time PCR. *Appl. Environ. Microbiol.* 75:965–969.
  69. Martín R, Langa S, Reviriego C, Jiménez E, Marín ML, Xaus J, Fernández L, Rodríguez JM. 2003. Human milk is a source of lactic acid bacteria for the infant gut. *J. Pediatr.* 143:754–758.
  70. Fernández A, Huang S, Seston S, Xing J, Hickey R, Criddle C, Tiedje J. 1999. How stable is stable? Function versus community composition. *Appl. Environ. Microbiol.* 65:3697–3704.
  71. Pérez-Chaia AP, Oliver G. 2003. Intestinal microflora and metabolic activity, p 77–98. *In* Fuller R, Perdigon G (ed), *Gut flora, nutrition, immunity and health*. Blackwell Publishing, Oxford, United Kingdom.
  72. Ortiz-Castro R, Contreras-Cornejo HA, Macías-Rodríguez L, López-Bucio J. 2009. The role of microbial signals in plant growth and development. *Plant Signal Behav.* 4:701–712.
  73. Phillips DA, Martínez-Romero E, Yang GP, Joseph JM. 2000. Release of nitrogen: a key trait in selecting bacterial endophytes for agronomically useful nitrogen fixation, p 205–217. *In* Ladha JK, Reddy PM (ed), *The quest for nitrogen fixation in rice*. IRRI, Manila, Philippines.
  74. Phillips DA, Joseph CM, Yang GP, Martínez-Romero E, Sanborn JR, Volpin H. 1999. Identification of lumichrome as a sinorhizobium enhancer of alfalfa root respiration and shoot growth. *Proc. Natl. Acad. Sci. U. S. A.* 96:12275–12280.
  75. Gibson GR, Cummings JH, Macfarlane GT. 1988. Use of a three-stage continuous culture system to study the effect of mucin on dissimilatory sulfate reduction and methanogenesis by mixed populations of human gut bacteria. *Appl. Environ. Microbiol.* 54:2750–2755.
  76. Ladha JK, Barraquillo WL, Watanabe I. 1983. Isolation and identification of nitrogen-fixing *Enterobacter cloacae* and *Klebsiella planticola* associated with rice plants. *Can. J. Microbiol.* 29:1301–1308.
  77. Madhaiyan M, Poonguzhali S, Kwon SW, Sa TM. 2009. *Methylophilus rhizosphaerae* sp. nov., a restricted facultative methylotroph isolated from rice rhizosphere soil. *Int. J. Syst. Evol. Microbiol.* 59:2904–2908.
  78. St-Pierre B, Wright AD. 27 April 2012, posting date. Diversity of gut methanogens in herbivorous animals. <http://dx.doi.org/10.1017/S1751731112000912>.
  79. Ormeño-Orrillo E, Hungria M, Martínez-Romero E. Dinitrogen-fixing prokaryotes. *In* Rosenberg E, DeLong EF, Stackebrandt E, Lory S,



- Thompson F (ed), The prokaryotes, vol 1. Symbiotic associations, biotechnology, applied microbiology, 4th ed, in press. Springer, New York, NY.
80. Behar A, Yuval B, Jurkevitch E. 2005. Enterobacteria-mediated nitrogen fixation in natural populations of the fruit fly *Ceratitis capitata*. *Mol. Ecol.* 14:2637–2643.
  81. Desai MS, Brunet A. 2012. Bacteroidales ectosymbionts of gut flagellates shape the nitrogen-fixing community in dry-wood termites. *ISME J.* 6:1302–1313.
  82. Ohkuma M. 2008. Symbioses of flagellates and prokaryotes in the gut of lower termites. *Trends Microbiol.* 16:345–352.
  83. Flint HJ, Bayer EA, Rincon MT, Lamed R, White BA. 2008. Polysaccharide utilization by gut bacteria: potential for new insights from genomic analysis. *Nat. Rev. Microbiol.* 6:121–131.
  84. Osawa R, Kuroiso K, Goto S, Shimizu A. 2000. Isolation of tannin-degrading lactobacilli from humans and fermented foods. *Appl. Environ. Microbiol.* 66:3093–3097.
  85. Gérard P, Lepercq P, Leclerc M, Gavini F, Raibaud P, Juste C. 2007. *Bacteroides* sp. strain D8, the first cholesterol-reducing bacterium isolated from human feces. *Appl. Environ. Microbiol.* 73:5742–5749.
  86. Derrien M, Collado MC, Ben-Amor K, Salminen S, de Vos WM. 2008. The mucin degrader *Akkermansia muciniphila* is an abundant resident of the human intestinal tract. *Appl. Environ. Microbiol.* 74:1646–1648.
  87. Calvaruso C, Turpault MP, Frey-Klett P. 2006. Root-associated bacteria contribute to mineral weathering and to mineral nutrition in trees: a budgeting analysis. *Appl. Environ. Microbiol.* 72:1258–1266.
  88. Puente ME, Bashan Y, Li CY, Lebsky VK. 2004. Microbial populations and activities in the rhizosphere of rock-weathering desert plants. I. Root colonization and weathering of igneous rocks. *Plant Biol.* 6:629–642.
  89. Soedarjo M, Hemscheidt TK, Borthakur D. 1994. Mimosine, a toxin present in leguminous trees (*Leucaena* spp.), induces a mimosine-degrading enzyme activity in some *Rhizobium* strains. *Appl. Environ. Microbiol.* 60:4268–4272.
  90. Allison MJ, Hammond AC, Jones RJ. 1990. Detection of ruminal bacteria that degrade toxic dihydroxypyridine compounds produced from mimosine. *Appl. Environ. Microbiol.* 56:590–594.
  91. Thomas F, Barbeyron T, Tonon T, Génicot S, Czjzek M, Michel G. 2012. Characterization of the first alginolytic operons in a marine bacterium: from their emergence in marine Flavobacteria to their independent transfers to marine Proteobacteria and human gut Bacteroides. *Environ. Microbiol.* 14:2379–2394.
  92. Kuiper I, Legendijk EL, Bloemberg GV, Lugtenberg BJ. 2004. Rhizoremediation: a beneficial plant-microbe interaction. *Mol. Plant Microbe Interact.* 17:6–15.
  93. Lugtenberg BJ, Kravchenko LV, Simons M. 1999. Tomato seed and root exudate sugars: composition, utilization by *Pseudomonas* biocontrol strains and role in rhizosphere colonization. *Environ. Microbiol.* 1:439–446.
  94. Haiser HJ, Turnbaugh PJ. 2012. Is it time for a metagenomic basis of therapeutics? *Science* 336:1253–1255.
  95. Mikov M. 1994. The metabolism of drugs by the gut flora. *Eur. J. Drug Metab. Pharmacokinet.* 19:201–207.
  96. Sousa T, Paterson R, Moore V, Carlsson A, Abrahamsson B, Basit AW. 2008. The gastrointestinal microbiota as a site for the biotransformation of drugs. *Int. J. Pharm.* 363:1–25.
  97. Pham TT, Tu Y, Sylvestre M. 2012. Remarkable ability of *Pandoraea pnomenus* B356 biphenyl dioxygenase to metabolize simple flavonoids. *Appl. Environ. Microbiol.* 78:3560–3570.
  98. Rodionov DA, Vitreschak AG, Mironov AA, Gelfand MS. 2003. Comparative genomics of the vitamin B12 metabolism and regulation in prokaryotes. *J. Biol. Chem.* 278:41148–41159.
  99. Albert MJ, Mathan VI, Baker SJ. 1980. Vitamin B12 synthesis by human small intestinal bacteria. *Nature* 283:781–782.
  100. Campbell GR, Taga ME, Mistry K, Lloret J, Anderson PJ, Roth JR, Walker GC. 2006. *Sinorhizobium meliloti* *bluB* is necessary for production of 5,6-dimethylbenzimidazole, the lower ligand of B12. *Proc. Natl. Acad. Sci. U. S. A.* 103:4634–4639.
  101. Morita H, Toh H, Fukuda S, Horikawa H, Oshima K, Suzuki T, Murakami M, Hisamatsu S, Kato Y, Takizawa T, Fukuoka H, Yshimura T, Itoh K, O'Sullivan D, McKay L, Ohno H, Kikuchi J, Masaoka T, Hattori M. 2008. Comparative genome analysis of *Lactobacillus reuteri* and *Lactobacillus fermentum* reveal a genomic island for reuterin and cobalamin production. *DNA Res.* 15:151–161.
  102. Ramotar K, Conly JM, Chubb H, Louie TJ. 1984. Production of menaquinones by intestinal anaerobes. *J. Infect. Dis.* 150:213–218.
  103. Shin SC, Kim SH, You H, Kim B, Kim AC, Lee KA, Yoon JH, Ryu JH, Lee WJ. 2011. *Drosophila* microbiome modulates host developmental and metabolic homeostasis via insulin signaling. *Science* 334:670–674.
  104. Bercik P, Denou E, Collins J, Jackson W, Lu J, Jury J, Deng Y, Blennerhasset P, Macri J, McCoy K, Verdu EF, Collins SM. 2011. The intestinal microbiota affect central levels of brain-derived neurotrophic factor and behavior in mice. *Gastroenterology* 141:599–609.
  105. Diaz Heijtz RD, Wang S, Anuar F, Qian Y, Björkholm B, Samuelsson A, Hibberd ML, Frossberg H, Pettersson S. 2011. Normal gut microbiota modulates brain development and behavior. *Proc. Natl. Acad. Sci. U. S. A.* 108:3047–3052.
  106. Kane M, Case LK, Kopaskie K, Kozlova A, MacDearmid C, Chervonisky AV, Golovkina TV. 2011. Successful transmission of a retrovirus depends on the commensal microbiota. *Science* 334:245–249.
  107. Jung SC, Martínez-Medina A, López-Raez JA, Pozo MJ. 2012. Mycorrhiza-induced resistance and priming of plant defenses. *J. Chem. Ecol.* 38:651–664.
  108. Saxelin M, Tynkkynen S, Mattila-Sandholm T, de Vos WM. 2005. Probiotic and other functional microbes: from markets to mechanisms. *Curr. Opin. Biotechnol.* 16:204–211.
  109. Puente ME, Li CY, Bashan Y. 2009. Rock-degrading endophytic bacteria in cacti. *Environ. Exp. Bot.* 66:389–401.
  110. Gibson GR, Rastall RA, Fuller R. 2003. The health benefits of probiotics and prebiotics, p 52–76. *In* Fuller R, Perdigón G (ed), Gut flora, nutrition, immunity and health. Blackwell Publishing, Oxford, United Kingdom.
  111. Rosenblueth M, Hynes MF, Martínez-Romero E. 1998. *Rhizobium tropici* *teu* genes involved in specific uptake of *Phaseolus vulgaris* bean-exudate compounds. *Mol. Genet.* 258:587–598.
  112. Tepfer D, Goldmann A, Pamboukdjian N, Maille M, Lepingle A, Chevalier D, Dénarié J, Rosenberg C. 1988. A plasmid of *Rhizobium meliloti* 41 encodes catabolism of two compounds from root exudate of *Calystegium sepium*. *J. Bacteriol.* 170:1153–1161.
  113. van Baaren L, Troost F, van der Meer C, Hooiveld G, Boekschoten M, Brummer RJM, Kleerebezem M. 2011. Human mucosal *in vivo* transcriptome responses to three lactobacilli indicate how probiotics may modulate human cellular pathways. *Proc. Natl. Acad. Sci. U. S. A.* 108(Suppl 1):4562–4569.
  114. Floch MH, Walker WA. 2008. Advances in clinical use of probiotics. *J. Clin. Gastroenterol.* 42:S45.
  115. Zimmer W, Stephan MP, Bothe H. 1984. Denitrification by *Azospirillum brasilense* Sp 7. *Arch. Microbiol.* 138:206–211.
  116. Delgado S, O'Sullivan E, Fitzgerald G, Mayo B. 2008. *In vitro* evaluation of the probiotic properties of human intestinal *Bifidobacterium* species and selection of new probiotic candidates. *J. Appl. Microbiol.* 104:1119–1127.
  117. Peix A, Velázquez E, Silva LR, Mateos PF. 2010. Key molecules involved in beneficial infection process in rhizobia-legume symbiosis, p 55–80. *In* Khan MH, Zaidi A, Musarrat J (ed), Microbes for legume improvement. Springer, Vienna, Austria.
  118. Downie JA. 2010. The roles of extracellular proteins, polysaccharides and signals in the interactions of rhizobia with legume roots. *FEMS Microbiol. Rev.* 34:150–170.
  119. González-Pasayo R, Martínez-Romero E. 2000. Multiresistance genes of *Rhizobium etli* CFN42. *Mol. Plant Microbe Interact.* 13:572–577.
  120. Ramachandran VK, East AK, Karunakaran R, Downie JA, Poole PS. 2011. Adaptation of *Rhizobium leguminosarum* to pea, alfalfa and sugar beet rhizospheres investigated by comparative transcriptomics. *Genome Biol.* 12:R106.
  121. López-Guerrero MG, Ormeño-Orrillo E, Acosta JL, Mendoza-Vargas A, Rogel MA, Ramírez MA, Rosenblueth M, Martínez-Romero J, Martínez-Romero E. 2012. Rhizobial extrachromosomal replicon variability, stability and expression in natural niches. *Plasmid* 68:149–158.
  122. Ramos-González MI, Campos MJ, Ramos JL. 2005. Analysis of *Pseudomonas putida* KT2440 gene expression in the maize rhizosphere: *in vitro* expression technology capture and identification of root-activated promoters. *J. Bacteriol.* 187:4033–4041.
  123. Karunakaran R, Ramachandran VK, Seaman JC, East AK, Mouhsine B. 2009. Transcriptomic analysis of rhizobium leguminosarum biovar

- viciae in symbiosis with host plants *Pisum sativum* and *Vicia cracca*. J. Bacteriol. 191:4002–4014.
124. Espinosa-Urgel M, Ramos JL. 2001. Expression of a *Pseudomonas putida* aminotransferase involved in lysine catabolism is induced in the rhizosphere. Appl. Environ. Microbiol. 67:5219–5224.
  125. Kim YC, Miller CD, Anderson AJ. 2000. Superoxide dismutase activity in *Pseudomonas putida* affects utilization of sugars and growth on root surfaces. Appl. Environ. Microbiol. 66:1460–1467.
  126. Langlois P, Bourassa S, Poirier GG, Beaulieu C. 2003. Identification of *Streptomyces coelicolor* proteins that are differentially expressed in the presence of plant material. Appl. Environ. Microbiol. 69:1884–1889.
  127. Begley M, Gahan CGM, Hill C. 2005. The interaction between bacteria and bile. FEMS Microbiol. Rev. 29:625–651.
  128. Gueimonde M, Garrigues C, van Sinderen D, de los Reyes-Gavilán CG, Margolles A. 2009. Bile-inducible efflux transporter from *Bifidobacterium longum* NCC2705, conferring bile resistance. Appl. Environ. Microbiol. 75:3153–3160.
  129. Ruiz L, Alvarez-Martín P, Mayo B, de los Reyes-Gavilán CG, Gueimonde M, Margolles A. 2012. Controlled gene expression in *Bifidobacteria* by use of a bile-responsive element. Appl. Environ. Microbiol. 78:581–585.
  130. Bron PA, Marco M, Hoffer SM, Van Mullekom E, de Vos MW, Kleerebezem M. 2004. Genetic characterization of the bile salt response in *Lactobacillus plantarum* and analysis of responsive promoters *in vitro* and *in situ* in the gastrointestinal tract. J. Bacteriol. 186:7829–7835.
  131. Ridlon JM, Kang DJ, Hylemon PB. 2006. Bile salt biotransformations by human intestinal bacteria. J. Lipid Res. 47:241–259.
  132. Blaut M, Schoefer L, Braune A. 2003. Transformation of flavonoids by intestinal microorganisms. Int. J. Vitam. Nutr. Res. 73:79–87.
  133. Cooper J. 2004. Multiple responses of rhizobia to flavonoids during legume root infection. Adv. Bot. Res. 41:1–62.
  134. Palumbo JD, Kado CI, Phillips DA. 1998. An isoflavonoid-inducible efflux pump in *Agrobacterium tumefaciens* is involved in competitive colonization of roots. J. Bacteriol. 180:3107–3113.
  135. Sims GK, Dunigan EP. 1984. Diurnal and seasonal variations in nitrogenase activity ( $C_2H_2$  reduction) of rice roots. Soil Biol. Biochem. 16:15–18.
  136. Hoogerwerf WA. 2006. Biologic clocks and the gut. Curr. Gastroenterol. Rep. 8:353–359.
  137. Costello EK, Gordon JI, Secor SM, Knight R. 2010. Postprandial remodeling of the gut microbiota in Burmese pythons. ISME J. 4:1375–1385.
  138. Comelli EM, Simmering R, Faure M, Donnicola D, Mansourian R, Rochat F, Corthesy-Theulaz I, Cherbut C. 2008. Multifaceted transcriptional regulation of the murine intestinal mucus layer by endogenous microbiota. Genomics 91:70–77.
  139. Hooper LV, Wong MH, Thelin A, Hansson L, Falk PG, Gordon JI. 2001. Molecular analysis of commensal host-microbial relationships in the intestine. Science 291:881–884.
  140. Rudrappa T, Czymmek KJ, Paré PW, Bais HP. 2008. Root-secreted malic acid recruits beneficial soil bacteria. Plant Physiol. 148:1547–1556.
  141. McNulty NP, Yatsunenko T, Hsiao A, Faith JJ, Muegge BD, Goodman AL, Henrissat B, Oozeer R, Cools-Portier S, Gobert G, Chervaux C, Knights D, Lozupone CA, Knight R, Duncan AE, Bain JR, Muehlbauer MJ, Newgard CB, Heath AC, Gordon JI. 2011. The impact of a consortium of fermented milk strains on the gut microbiome of gnotobiotic mice and monozygotic twins. Sci. Transl. Med. 3:106.
  142. Macpherson AJ, Harris NL. 2004. Interactions between commensal intestinal bacteria and the immune system. Nat. Rev. Immunol. 4:478–485.
  143. Farag MA, Ryu CM, Sumner LW, Paré PW. 2006. GC-MS SPME profiling of rhizobacterial volatiles reveals prospective inducers of growth promotion and induced systemic resistance in plants. Phytochemistry 67:2262–2268.
  144. Mathesius M, Mulders S, Gao M, Teplitski M, Caetano-Anollés G, Rolfe BG, Bauer WD. 2003. Extensive and specific responses of a eukaryote to bacterial quorum-sensing signals. Proc. Natl. Acad. Sci. U. S. A. 100:1444–1449.
  145. Gao M, Teplitski M, Robinson JB, Bauer WD. 2003. Production of substances by *Medicago truncatula* that affect bacterial quorum sensing. Mol. Plant Microbe Interact. 16:827–834.
  146. Teplitski M, Mathesius U, Rumbaugh KP. 2011. Perception and degradation of *N*-acyl homoserine lactone quorum sensing signals by mammalian and plant cells. Chem. Rev. 111:100–116.
  147. Ortiz-Castro R, Díaz-Pérez C, Martínez-Trujillo M, del Río RE, Campos-García J, López-Bucio J. 2011. Transkingdom signaling based on bacterial cyclodipeptides with auxin activity in plants. Proc. Natl. Acad. Sci. U. S. A. 108:7253–7258.
  148. Oldroyd GE, Murray JD, Poole PS, Downie JA. 2011. The rules of engagement in the legume-rhizobial symbiosis. Annu. Rev. Genet. 45:119–144.
  149. Ryu JH, Kim SH, Lee HY, Bai JY, Nam YD, Bae JW, Lee DG, Shin SC, Ha EM, Lee WJ. 2008. Innate immune homeostasis by the homeobox gene *caudal* and commensal-gut mutualism in *Drosophila*. Science 319:777–782.
  150. Kim EK, Kim SH, Nam HJ, Choi MK, Lee KA, Choi SH, Seo YY, You H, Kim B, Lee WJ. 2012b. Draft genome sequence of *Commensalibacter intestini* A911T, a symbiotic bacterium isolated from *Drosophila melanogaster* intestine. J. Bacteriol. 194:1246.
  151. Roh SW, Nam YD, Chang HW, Kim KH, Kim MS, Ryu JH, Kim SH, Lee WJ, Bae JW. 2008. Phylogenetic characterization of two novel commensal bacteria involved with innate immune homeostasis in *Drosophila melanogaster*. Appl. Environ. Microbiol. 74:6171–6177.
  152. Ormeño-Orrillo E, Rosenblueth M, Luyten E, Vanderleyden J, Martínez-Romero E. 2008. Mutations in lipopolysaccharide biosynthetic genes impair maize rhizosphere and root colonization of *Rhizobium tropici* CIAT899. Environ. Microbiol. 10:1271–1284.
  153. Bang C, Schilhabel A, Weidenbach K, Kopp A, Goldmann T, Gutschmann T, Schmitz RA. 2012. Effects of antimicrobial peptides on methanogenic archaea. Antimicrob. Agents Chemother. 56:4123–4130.
  154. Lee WJ. 2009. Bacterial-modulated host immunity and stem cell activation for gut homeostasis. Genes Dev. 23:2260–2265.
  155. Round JL, Mazmanian SK. 2009. The gut microbiota shapes intestinal immune responses during health and disease. Nat. Rev. Immunol. 9:313–323.
  156. Peumans WJ, Van Damme EJ. 1995. Lectins as plant defense proteins. Plant Physiol. 109:347–352.
  157. Vaishnava S, Yamamoto M, Severson KM, Ruhn KA, Yu X, Koren O, Ley R, Wakeland EK, Hooper LV. 2011. The antibacterial lectin RegIII $\alpha$  promotes the spatial segregation of microbiota and host in the intestine. Science 334:255–258.
  158. Gibson GR, Wang X. 1994. Regulatory effects of bifidobacteria on the growth of other colonic bacteria. J. Appl. Microbiol. 77:412–420.
  159. Kreth J, Merritt J, Qi F. 2009. Bacterial and host interactions of oral streptococci. DNA Cell Biol. 28:397–403.
  160. Edwards A, Frederix M, Wisniewski-Dyé F, Jones J, Zorreguieta A, Downie JA. 2009. The *cin* and *rai* quorum-sensing regulatory systems in *Rhizobium leguminosarum* are coordinated by ExpR and CinS, a small regulatory protein coexpressed with CinI. J. Bacteriol. 191:3059–3067.
  161. Jafra S, Przysocka J, Czajkowski R, Michta A, Garbeva P, Van der Wolf JM. 2006. Detection and characterization of bacteria from the potato rhizosphere degrading *N*-acyl-homoserine lactone. Can. J. Microbiol. 52:1006–1015.
  162. Kroer N, Barkay T, Sorensen S, Weber D. 1998. Effect of root exudates and bacterial metabolic activity on conjugal gene transfer in the rhizosphere of a marsh plant. FEMS Microbiol. Ecol. 25:375–384.
  163. Shoemaker NB, Vlamakis H, Hayes K, Salyers AA. 2001. Evidence for extensive resistance gene transfer among *Bacteroides* spp. and among *Bacteroides* and other genera in the human colon. Appl. Environ. Microbiol. 67:561–568.
  164. Arias MC, Danchin EGJ, Coutinho PM, Henrissat B, Ball S. 2012. Eukaryote to gut bacteria transfer of a glycoside hydrolase gene essential for starch breakdown in plants. Mob. Genet. Elements 2:81–87.
  165. Stern A, Mick E, Tirosh I, Sagy O, Sorek R. 2012. CRISPR targeting reveals a reservoir of common phages associated with the human gut microbiome. Genome Res. 22:1985–1994.
  166. Sullivan JT, Ronson CW. 1998. Evolution of rhizobia by acquisition of a 500-kb symbiosis island that integrates into a *phe*-tRNA gene. Proc. Natl. Acad. Sci. U. S. A. 95:5145–5149.
  167. Husník F, Chrudimský T, Hypša V. 2011. Multiple origins of endosymbiosis within the Enterobacteriaceae ( $\gamma$ -Proteobacteria): convergence of complex phylogenetic approaches. BMC Biol. 9:87.
  168. Fukatsu T, Hosokawa T. 2002. Capsule-transmitted gut symbiotic bac-

- terium of the Japanese common plataspid stinkbug, *Megacopta punctatissima*. *Appl. Environ. Microbiol.* **68**:389–396.
169. López-López A, Rosenblueth R, Martínez J, Martínez-Romero E. 2010. Rhizobial symbioses in tropical legumes and non-legumes. *Soil Biol.* **21**: 163–184.
170. McCutcheon JP, Moran NA. 2012. Extreme genome reduction in symbiotic bacteria. *Nat. Rev. Microbiol.* **10**:13–26.
171. de Bruijn FJ. 2013. Molecular microbial ecology of the rhizosphere, vol I and II. Wiley-Blackwell, Hoboken, New Jersey.
172. Dessaux Y, Hinsinger P, Lemanceau P. 2007. Rhizosphere: achievements and challenges. Springer Science Press, Berlin, Germany.
173. Fuller R, Perdigon G. 2003. Gut flora, nutrition, immunity and health. Blackwell Publishing, Oxford, United Kingdom.
174. Pinto R, Varanini Z, Nannipieri P. 2007. The rhizosphere: biochemistry and organic substances at the soil-plant interface. Taylor and Francis Group/CRC Press, Boca Raton, FL.
175. Sadowsky MJ, Whitman RL. 2011. The fecal bacteria. ASM Press, Washington, DC.