Characteristics of the leaf parts of some a traditional Korean salad plants used for food



Sang-Uk Chon,¹ Buk-Gu Heo,² Yong-Seo Park,³ Ja-Yong Cho⁴ and Shela Gorinstein^{5*}

¹Callus Co. Ltd, Gwangju Institute of Science and Technology, Gwangju 500-712, South Korea

²Naju Foundation of Natural Dyeing Culture, Naju 520-931, South Korea

³Department of Horticultural Science, Mokpo National University, Muan 534-729, South Korea

⁴Department of Medicinal Resources and Horticulture Development, Jeonnam Provincial College, Damyang 517-802, South Korea

⁵Department of Medicinal Chemistry and Natural Products, School of Pharmacy, The Hebrew University – Hadassah Medical School, Jerusalem, Israel

Abstract

BACKGROUND: Total phenolics content, antioxidant activity and cytotoxicity of the methanol extracts from leaf parts of 13 Korean traditional salad plants were investigated in order to determine their properties.

RESULTS: The highest phenolics content (mg ferulic acid equivalents kg⁻¹ dry weight (d.w.), omit one) was found in methanol extracts from *Polygonum aviculare*, at 293.7 ± 6.0, followed by *Euonymus alatus*, at 250.7 ± 3.3, *Saxifraga stolonifera*, at 125.0 ± 8.1 and *Ligularia fischeri*, at 122.5 ± 5.9. The methanol plant extracts dose-dependently increased free radical scavenging activity. Methanol extracts of *Polygonum aviculare*, *Euonymus alatus* and *Saxifraga stolonifera*, at 31 mg kg⁻¹, exhibited the highest 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity (%) by 90.8 ± 4.2, 85.7 ± 3.9 and 64.1 ± 3.2, respectively. According to 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay, the methanol extracts from *Portulaca oleracea* (IC₅₀ < 25.0 µg mL⁻¹) showed the highest cytotoxicity against Calu-6, followed by *Plantago asiatica* (49.2 µg mL⁻¹) and *Osmunda japonica* (89.6 µg mL⁻¹).

CONCLUSION: Total phenolics content of the tested plant extracts was correlated with the DPPH radical scavenging activity, suggesting the phenolics compounds are contributing to the antioxidant properties of Korean salad plants. The leaf parts of the 13 Korean traditional salad plants described here that are currently used as foods may also provide some benefit to human health, and research into their potential benefits as preventative and/or therapeutic agents is warranted.

© 2008 Society of Chemical Industry

Keywords: Korean salad plants; methanol extracts; total phenolics; radical scavenging activity; cytotoxicity

INTRODUCTION

Some plants exhibit various bioactivities, including antioxidant, anti-inflammatory, anticancer and antidiabetic.^{1–3} Recently, there has been a worldwide trend towards the use of wild plants due to their bioactive phytochemicals, in particular, phenolics.^{4,5} Phenolic compounds are secondary metabolites that are synthesized by plants during normal development and in response to stress conditions such as infection, wounding and UV radiation.^{3–5} These compounds occur ubiquitously in plants and are a diversified group of phytochemicals derived from phenylalanine and tyrosine.^{6,7}

Free radical scavenging is generally the accepted mechanism for antioxidants inhibiting lipid oxidation. Antioxidants, inhibitors of lipid peroxidation, are important not only for food protection but also for the defense of living cells against oxidative damage.^{8,9} Therefore, not only antioxidants of natural products, but also synthesized antioxidants are widely used.¹⁰⁻¹² The toxic and other unfavorable effects of synthesized food antioxidants have been widely noted.13,14 Phenolic compounds, such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), and tertbutylhydroquinone (TBHQ), have been widely used as synthetic antioxidants in food. These substances are considered as safe natural antioxidants; however, they do not always provide effective protection against oxidation in vitro.15 Nevertheless, these phenolic antioxidants are still used extensively as food antioxidants because of their low cost. However, it must be mentioned that as early as in 1975 it was

E-mail: gorin@cc.huji.ac.il

^{*} Correspondence to: Shela Gorinstein, Department of Medicinal Chemistry and Natural Products, School of Pharmacy, The Hebrew University – Hadassah Medical School, Jerusalem 91120, Israel

⁽Received 9 September 2007; revised version received 5 February 2008; accepted 12 March 2008) Published online 9 July 2008; DOI: 10.1002/jsfa.3304

^{© 2008} Society of Chemical Industry. J Sci Food Agric 0022–5142/2008/\$30.00

reported that administration of these phenolic antioxidants in doses of 50 mg kg⁻¹ per day to rodents and monkeys led to certain pathological changes, including enzyme and lipid alterations as well as carcinogenic effects.¹⁶ Therefore, research on new natural antioxidants has gained momentum as they are considered, rightly or wrongly, not to pose any health risk to consumers.^{17,18}

Antioxidants in plants include flavonoids, phenolic acids, lignan precursors, terpenes, mixed tocopherols, phospholipids and polyfunctional organic acids, and also plant extracts such as those of rosemary and sage.^{18,19} It was reported that the antioxidant activity of the antioxidants mentioned is in the following order: rosmarinic acid > caffeic acid phenethyl ester > caffeic acid > chlorogenic acid > tocopherol > ferulic acid > ferulic acid phenethyl ester > BHT.²⁰

Vegetables, fruits and whole grains contain a wide variety of phytochemicals that have the potential to interfere with the development of cancer.^{1–3} These phytochemicals are isothiocyanates (cruciferous vegetables), carotenoids, including α -carotene, γ -carotene, β -cryptoxanthin, zeaxanthin(e), lutein, lycopene (tomatoes), resveratrol (grapes and wine), ellagic acid (various berries), glutathione-S-transferase (garlic), diallyl sulfide (garlic), genestine (soybean), curcumine (turmeric), indole-3-carbinol, inositol, organosulfur compounds, sulforaphane, squalene, and terpenes.²¹ Also regular consumption of tea decreased the risk of various types of cancer.^{22,23}

The Korean medicinal plants that are investigated in this study have been used for a long time as traditional seasoned salads, and were screened for bioactive effects of these functional foods.²⁴ Earlier studies showed that extracts from *Areca catechu* var. *dulcissima* possess antidepressant properties.²⁵ Au and Lam²⁶ suggested that the methanol extracts of *Paeonia suffruticosa* potently inhibit human immunodeficiency virus (HIV)-1 integrase. Lee *el al.*²⁷ reported that silymarin and silybin purified from *Silybum marianum* have the potential to inhibit activities against oxidation of ¹²⁵I-LDL by macrophages and endothelial cells. Therefore these Korean plants, used as salad, could act as preventative or therapeutic agents.

The objective of this research was to determine total phenolics content of the methanol extracts from leaf parts of the 13 Korean salad plants, their antioxidant activity and cytotoxicity. In order to receive reliable data the following were used: (a) for determination of total phenolics content: the Folin–Ciocalteu assay; (b) for assessment of the radical scavenging activity: 1,1-diphenyl-2-picrylhydrazyl (DPPH) assay, because of its ability in a relatively short time compared to other methods to evaluate antioxidative activities;²⁸ (c) for anticancer activity: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay.

As far as we know, there are no published results of similar investigations.

MATERIALS AND METHODS Chemicals

Folin-Ciocalteu reagent, MTT, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT) and ascorbic acid were obtained from Sigma Chemical Co. (St Louis, MO, USA). All reagents were of analytical grade. Deionized and distilled water were used throughout. The cell lines were purchased from Korean Cell Line Bank (KCLB).

Plant material

Leaf parts of 13 Korean medicinal salad plants (Saxifraga stolonifera (L.) Meeb, Pteridium aquilinum, Hemerocallis minor, Plantago asiatica, Osmunda japonica, Portulaca oleracea, Synurus deltoides, Ligularia fischeri, Rumex acetosa, Polygonum aviculare, Symplocarpus renifolius, Chenopodium album, and Euonymus alatus) grown in a mountain area of Suncheon City, Korea, were harvested at a vegetative stage in June 2006. The samples were immediately freeze-dried at -40 °C for 5 days, ground with a Wiley mill to pass a 1 mm screen, and stored in a refrigerator at 2 °C until used.

The samples were extracted three times with 95% methanol in a shaker at 24 h intervals for 3 days at room temperature. The extracts were then filtered through a Whatman No. 1 filter paper. The collected filtrate was evaporated to dryness under vacuum at $40 \,^{\circ}$ C using a rotary evaporator (N-1000V-W, Eyela, Japan). After evaporation, the yield of dried methanol extract was about 10% of the original plant sample. The methanol extracts from each plant were used for determination of total phenolic content, DPPH radical scavenging activity, and cytotoxicity.

Total phenolics content

The content of total phenolics (TP) was measured using the classical Folin-Ciocalteu assay.²⁹ 5 mL of (Thermo Scientific Barnstead NaNOpure[™] Water Purification System, Analytical, EW-99277-00) Nanopure water, 0.5–1.0 mL of sample, and 1.0 mL of Folin-Ciocalteu reagent were added to a 25 mL volumetric flask. The above-mentioned substances were mixed and allowed to stand for 5-8 min at room temperature. Next, 10 mL of a 7% sodium carbonate solution was added, followed by the addition of Nanopure water filled to volume. Solutions were mixed and allowed to stand at room temperature for 2h. Sample aliquots were filtered through a Whatman 0.45 m poly(tetrafluoroethylene) filter prior to the determination of TP content using a UV-1650 spectrophotometer (Shimadzu, Kyoto, Japan) monitoring at 640 nm. TP were standardized against ferulic acid and expressed as ppm of ferulic acid equivalents (FAE). The linearity range for this assay was determined as $0.5-5.0 \text{ mg L}^{-1}$ FAE ($R^2 = 0.9990$), giving an absorbance range of 0.050–0.555 AU.

Radical scavenging activity

Free radical scavenging activity of the methanol extracts was determined using the classical DPPH

method. Each methanol extract at various concentrations (31, 63, 125, 250, and 500 mg kg⁻¹) was added to a $1.5 \times 10^{-4} \text{ mol } \text{L}^{-1}$ solution of DPPH in methanol and the reaction mixture was shaken vigorously. The amount of DPPH remaining was determined at 520 nm, and the radical scavenging activity was obtained from the following equation: radical scavenging activity (%) = $\{(OD_{control} -$ $OD_{sample})/OD_{control}$ × 100. The antioxidant activity of plant extracts was partially expressed as IC₅₀, which was defined as the concentration $(mg kg^{-1})$ of extract required to inhibit the formation of DPPH radicals by 50%.

Anticancer activity

Anticancer activity of methanol extracts of the studied plants on human cancer cell lines (Calu-6 for human pulmonary carcinoma and SNU-601 for human gastric carcinoma) were measured using the MTT assay. Cell lines for the MTT assay were purchased from KCLB. Cells were grown in RPMI-1640 medium at 37 °C under 5% CO₂ in a humidified incubator. Cells were harvested, counted $(3 \times 10^4 \text{ cells mL}^{-1})$, and transferred to a 96-well plate, and incubated for 24 h prior to the addition of test compounds. Serial dilutions of test samples were prepared by dissolving compounds in dimethyl sulfoxide (DMSO), followed by dilution with RPMI-1640 medium to give final concentrations of 25, 50, 100, 200, 400, and $800 \,\mu g \,m L^{-1}$. Stock solutions of samples were prepared for cell lines at 90μ L and samples at 10μ L, and incubated for 72 h. MTT solution at 5 mg mL^{-1} was dissolved in 1 mL of phosphate buffer solution (PBS), and $10\,\mu\text{L}$ of this was added to each of the 96 wells.³⁰ The wells were wrapped with aluminium foil and incubated at 37 °C for 4 h. The solution in each well containing media, unbound MTT and dead cells was removed by suction and 150 µL of DMSO was added to each well. The plates were then shaken and optical density was recorded using a micro-plate reader at 540 nm. Distilled water was used as positive control and DMSO as solvent control. Controls and samples were assayed in duplicate for each concentration and replicated three times for each cell line. Cytotoxicity was established by comparing the absorbance between the samples and the control. The values were then used to iteratively calculate the concentration of plant extracts required to cause a 50% reduction (IC₅₀) in growth (cell number) for each cell line.

Statistics

To verify the statistical significance, mean \pm SD of three independent measurements were calculated. Differences between groups were tested by twoway ANOVA. In the assessment of the antioxidant potential, Spearman correlation coefficients (R) were used. Linear regressions were also calculated. P-values of <0.05 were considered significant.

RESULTS AND DISCUSSION Total phenolics content

The amounts of TP in methanol extracts (mg FAE kg^{-1} d.w.) for Polygonum aviculare and Euonymus alatus were 293.7 ± 6.0 and 250.7 ± 3.3 , respectively, and were significantly higher (P < 0.05) than in Saxifraga stolonifera and Ligularia fischeri (125.0 \pm 8.1 and 122.5 ± 5.9 , respectively). The lowest content of TP was in the extracts of Chenopodium album, Hemerocallis minor, and Symplocarbus renifolius (P < 0.05 in all three cases) (Fig. 1). The result was highly consistent with the finding of DPPH radical scavenging activity.³¹ Also Zhou and Yu³² reported that TP content of the tested vegetable extracts was correlated with DPPH radical scavenging activity, suggesting that TP can play a major role in the antioxidant activity of plant materials.

DPPH radical scavenging activity

Table 1 summarizes the results of the determination of DPPH radical scavenging activity. As can bee seen, methanol extracts of Polygonum aviculare had the highest DPPH radical scavenging activity, followed by Euonymus alatus and Saxifraga stolonifera, indicating IC_{50} values below 31.0 mg kg⁻¹. These values of Polygonum aviculare and Euonymus alatus showed higher activity than those of synthetic antioxidants (vitamin C and BHT), with IC₅₀ values of <31.0 and 113.0 mg kg^{-1} , respectively. Methanol extracts of Polygonum aviculare, Euonymus alatus, and Saxifraga stolonifera at 31.0 mg kg⁻¹ exhibited the highest DPPH radical scavenging activity (%) by $90.8 \pm 4.2, 85.7 \pm$ 3.9, and 64.1 ± 3.2 , respectively.

DPPH radical scavenging activity data for Chenopodium album extract was the lowest (P < 0.05). All samples of plant species showed that DPPH radical scavenging activity is dose-dependent. The results

350

300

250

200

Content of total phenolics (mg kg⁻¹) 150 100 50 0 Saxifraga s. Pteridium a. Hemerocallis m. Plantago a. Portulaca o. Synurus d. igularia f. Rumex a. Polygonum a. Symplocarpus r. Chenopodium a. Euonymus a. Osmunda Korean salad plants Figure 1. Total phenolic amount (mg FAE kg⁻¹ dry weight) of

methanol extracts from the leaf parts of 13 Korean medicinal salad plants.

	Extract concentration (mg kg $^{-1}$)					
Scientific name	31	63	125	250	500	IC_{50}^{\dagger}
Saxifraga stolonifera	$64.1 \pm 3.2g$	$90.4 \pm 4.2h$	$91.1 \pm 4.2h$	$91.1 \pm 4.2h$	$91.6 \pm 4.2h$	<31
Pteridium aquilinum	$15.6 \pm 1.1c$	$25.8 \pm 1.8 d$	$45.5 \pm 2.3c$	$77.2 \pm 3.5 d$	$86.4 \pm 4.0c$	140
Hemerocallis minor	$9.8 \pm 0.7 b$	$14.8 \pm 1.0b$	21.7 ± 1.8a	$37.8 \pm 2.4b$	$65.8 \pm 3.0b$	360
Plantago asiatica	$20.5 \pm 1.6c$	$31.6 \pm 1.9e$	$56.9 \pm 2.9e$	$88.0 \pm 4.0h$	$90.4 \pm 4.1h$	106
Osmunda japonica	$25.2 \pm 1.8d$	$44.5 \pm 2.1 f$	$74.7 \pm 3.3h$	$88.7 \pm 4.0h$	$87.8 \pm 3.9c$	75
Portulaca oleracea	4.9 ± 0.3a	9.9 ± 0.7a	19.8 ± 1.6a	$37.2 \pm 2.2a$	$67.2 \pm 3.1 b$	361
Synurus deltoides	$11.3 \pm 0.8b$	$19.5 \pm 1.6c$	$34.2 \pm 2.1b$	$60.3 \pm 3.1c$	$85.4 \pm 3.9c$	200
Ligularia fischeri	$37.2 \pm 2.2e$	$64.8 \pm 3.2g$	$89.4 \pm 4.1 h$	$90.5 \pm 4.1h$	$90.6 \pm 4.1 h$	46
Rumex acetosa	$49.1 \pm 2.5 f$	$75.7 \pm 3.3h$	$89.5 \pm 4.1 h$	$90.0 \pm 4.1h$	$90.7 \pm 4.1h$	32
Polygonum aviculare	90.8 ± 4.2a	$91.4 \pm 4.2h$	$91.4 \pm 4.2h$	$91.8 \pm 4.2h$	$92.8 \pm 4.3h$	<31
Symplocarpus renifolius	$9.8 \pm 0.7 b$	$16.9 \pm 1.3b$	$30.8 \pm 2.0b$	$51.1 \pm 2.5c$	$79.7 \pm 3.6c$	242
Chenopodium album	0	0	0	0	4.1 ± 0.3a	-
Euonymus alatus	$85.7 \pm 3.9 h$	$91.2 \pm 4.2h$	$91.2 \pm 4.2h$	$91.2 \pm 4.2h$	$91.2 \pm 4.2h$	<31
Ascorbic acid	$81.8 \pm 3.7h$	$96.1 \pm 4.5h$	$96.1 \pm 4.5h$	$96.7 \pm 4.5h$	$96.9 \pm 4.5h$	<31
BHT	$15.6\pm1.1c$	$33.5\pm2.1e$	$55.2\pm2.8g$	$81.3\pm3.7h$	$92.4\pm4.3h$	113

Table 1. DPPH radical scavenging activity in methanol extracts from the leaf parts of 13 Korean traditional salad plants in comparison with synthetic antioxidants

Values are means \pm SD of three measurements. Means in columns without letters in common differ significantly (P < 0.05).

1,1-diphenyl-2-picrylhydrazyl (DPPH); butylated hydroxytoluene (BHT).

[†] Extract concentrations, which show 50% activity of DPPH radical scavenging, were determined by interpolation.

show that antioxidant activity compounds could be of different amounts in various plant species. The results of this study suggest that Ixeris dentate and Aster scaber are to be preferred against synthetic antioxidants. Also others have reported that antioxidant activity of plants is higher than that of synthetic antioxidants.³³ Those authors investigated methanol extracts of nine medicinal plants traditionally used in Chinese medicine versus resveratrol. They found relatively high levels of DPPH radical scavenging activity in extracts of Areca catechu var. dulcissima, Paeonia suffruticosa and Cinnamomun cassia ($IC_{50} < 6.0 \,\mu g \,m L^{-1}$). The extracts of Areca catechu var. dulcissima showed higher antioxidant activity than resveratrol in all experiments. In some reports³⁴⁻³⁶ solvents were applied with different polarity for polyphenols extraction and these solvents had a significant effect on polyphenol content and antioxidant activity. Others have shown that polyphenols and antioxidant capacities in water extracts were higher than in methanol. The correlation coefficients between polyphenols and antioxidant capacities of Prolipid with 1,1-diphenyl-2-picrylhydrazyl radical assay were about 0.97.34 It was also shown that 50% of ethanol extract from black herbal mate tea had the greatest antioxidant activity.34-36 The methanol extract of Ulmus davidiana³⁶ exhibited strong antioxidant activity in the tested model systems. U. davidiana extracts may be exploited as biopreservatives in food applications as well as for health supplements of functional food, to alleviate oxidative stress. Our results also showed high phenolics content and antioxidant activity in the methanol extracts from *Polygonum aviculare*, Euonymus alatus, and Saxifraga stolonifera. However, extracts of the plants showed low anticancer activity. The correlation coefficient between the extracted polyphenols with ethanol and antioxidants was about 0.88 (Fig. 2). The relatively high correlation between the above-mentioned variables in the studied extracts was compared with tea, Prolipid and *U. davidiana* extracts.^{34–36} Results of this investigation showed that traditional Korean salad plants used for food exercise both antioxidant and anticancer activities. However, the levels of the antioxidant and anticancer activities in some of the studied plants were different.

Cytotoxic effect of methanol extracts

The results of the cytotoxic effect of methanol extracts from the leaf parts of 13 Korean salad plants on two human cancer cell lines (Calu-6 and SNU-601) are summarized in Table 2. *Portulaca oleracea* exhibited the greatest influence on the Calu-6 human pulmonary carcinoma cell line ($<25.0 \pm 1.7$), followed by *Plantago asiatica* (49.2 ± 4.2). *Osmunda japonica* had the greatest effect on SNU-601 human gastric carcinoma cell line (152.9 ± 8.1), followed by *Portulaca oleracea* (213.9 ± 10.1). All these extracts



Figure 2. Correlation between amount of polyphenols (mg FAE kg^{-1} dry weight) and antioxidant activity (%) in Korean salad plants.

Table 2. Cytotoxic effect of methanol extracts from the leaf parts of
13 Korean salad plants on two human cancer cell lines

	$IC50^{a}$ ($\mu g m L^{-1}$)			
Scientific name	Calu-6 ^b	SNU-601 ^b		
Saxifraga stolonifera	$449.2 \pm 19.1 g$	$471.2\pm19.1f$		
Pteridium aquilinum	$220.1 \pm 10.2d$	$323.8 \pm 12.2d$		
Hemerocallis minor	$181.4 \pm 9.1c$	$344.0 \pm 13.3 d$		
Plantago asiatica	$49.2 \pm 4.2a$	$276.6 \pm 11.8c$		
Osmunda japonica	$89.6 \pm 6.1 b$	$152.9 \pm 8.11a$		
Portulaca oleracea	<25.0 ± 1.7a	$213.9 \pm 10.1 b$		
Synurus deltoides	$193.6 \pm 9.2c$	313.8 ± 12.3a		
Ligularia fischeri	$195.9 \pm 9.3c$	$283.7 \pm 12.0c$		
Rumex acetosa	$344.0 \pm 12.3 f$	$424.8 \pm 19.0e$		
Polygonum aviculare	$752.4 \pm 23.1 h$	$447.1 \pm 19.1e$		
Symplocarpus renifolius	$188.7 \pm 10.3c$	$335.8 \pm 12.3 d$		
Chenopodium album	$378.4 \pm 13.2 f$	$339.8 \pm 12.3 d$		
Euonymus alatus	$297.2\pm12.1\mathrm{e}$	$412.5\pm18.9\text{e}$		

Values are means \pm SD of three measurements. Means in columns without letters in common differ significantly (P < 0.05).

^a Extract concentrations, which inhibit 50% growth of cells, were determined by interpolation.

^b Calu-6 is human pulmonary carcinoma and SNU-601 is human gastric carcinoma.

were tested for their antiproliferative activity on tumor cell lines Calu-6 and SNU-601 by the MTT assay.³⁰ A dose-dependent inhibition of cell proliferation was observed for most of the methanol extracts tested in this study. It was recorded that the methanol extracts exhibited more inhibition on Calu-6 cell line than of SNU-601. Methanol extracts at $200 \,\mu g \,m L^{-1}$ from Portulaca oleracea exhibited the highest anticancer activity on Calu-6 and SNU-601 tumor cell lines, by 100% and 60%, respectively, while the methanol extracts from Polygonum aviculare at the same concentration exhibited the lowest activity, by 32% and 60%, respectively (Fig. 3). These results, however, were not consistent with the findings of DPPH radical scavenging activity or total phenolic content. Methanol extracts from *Portulaca oleracea* (IC₅₀ < $25.0 \,\mu g \,m L^{-1}$) showed the most potent cytotoxicity on Calu-6 cell line, followed by *Plantago asiatica* $(49.2 \,\mu g \,m L^{-1})$, Osmunda japonica (89.6 μ g mL⁻¹), and Hemerocallis *minor* $(181.4 \,\mu g \,m L^{-1})$. However, cytotoxicity of the methanol extracts from all plants against SNU-601 was much lower than that against Calu-6 cell line. Methanol extracts from Osmunda japonica with IC_{50} of $152.9 \,\mu g \,m L^{-1}$ showed the highest activity against SNU-601, followed by Portulaca oleracea $(213.9 \,\mu\text{g}\,\text{mL}^{-1})$, Plantago asiatica $(276.6 \,\mu\text{g}\,\text{mL}^{-1})$ and Ligularia fischeri (283.7 μ g mL⁻¹). Also Manosroi et al.³⁷ reported the antiproliferative activity of essential oil extracted from 17 Thai medicinal plants on human mouth epidermal carcinoma (KB) and murine leukemia (P338) cell lines using MTT assay. They found that guava (Psidium guajava L.) leaf and sweet basil oils exhibited the highest antiproliferative activity in KB and P388 cell lines, respectively.



Figure 3. Cytotoxic effect of methanol extracts from the leaf parts of *Portulaca oleracea* (A) and *Polygonum aviculare* (B) on human cancer cell lines, Calu-6 and SNU-601.

CONCLUSION

This investigation shows that the content of total phenolics in 13 Korean traditional salad plants is high: the significantly highest contents were found in Polygonum aviculare and Euonymus alatus. In addition, methanol extracts of the same plants were characterized by the highest DPPH radical scavenging activity. The salad plants dose-dependently increased free radical scavenging activity. The TP level is highly correlated with the free radical scavenging activity. The highest influence on the Calu-6 human pulmonary carcinoma cell line was found in the methanol extract of Portulaca oleracea, followed by that of Plantago asiatica, while extracts of Osmunda japonica and Portulaca oleracea had the highest impact on SNU-601 human gastric carcinoma cell line. Therefore, leaf parts of Korean traditional salad plants could be recommended as preventative or/and therapeutic agents mainly for atherosclerotic heart disease in addition to appropriate prescription drugs.

ACKNOWLEDGEMENTS

This research was conducted with support from the 2005 ARPC research fund (105088-33-2-HD110). Appreciation is expressed to Dr Sook-Young Lee at Chonsun University, Gwangju, Korea, for her technical assistance.

REFERENCES

- 1 Cai Y, Luo O, Sun M and Corke H, Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sci* 74:2157–2184 (2004).
- 2 Cho WCS and Leung KN, In vitro and in vivo anti-tumor effects of Astragalus membranaceus. Cancer Lett 252:43-54 (2007).
- 3 Russo GL, Ins and outs of dietary phytochemicals in cancer chemoprevention. *Biochem Pharmacol* 74:533–544 (2007).
- 4 Canter PH, Thomas H and Ernst E, Bringing medicinal plants into cultivation: opportunities and challenges for biotechnology. *Trends Biotechnol* 23:180–185 (2005).
- 5 Estomba D, Ladio A and Lozada M, Medicinal wild plant knowledge and gathering patterns in a Mapuche community from North-western Patagonia. *J Ethnopharmacol* 103:109-119 (2006).
- 6 Harborne JB and Turner M, *Plant Chemosystematics*. Academic Press, London (1984).
- 7 Shahidi F and Naczk M, *Phenolics in Food and Nutraceuticals:* Sources, Applications and Health Effects. CRC Press. Boca Raton, FL (2004).
- 8 Chen L, Haught WH, Yang B, Saldeen TG, Parathasarathy S and Mehta J, Preservation of endogenous antioxidant activity and inhibition of lipid peroxidation as common mechanisms of antiatherosclerotic effects of vitamin E, lovastatin and amlodipine. *J Am Coll Cardiol* **30**:569–575 (1997).
- 9 Mastaloudis A, Morrow JD, Hopkins DW, Devaraj S and Traber MG, Antioxidant supplementation prevents exerciseinduced lipid peroxidation, but not inflammation, in ultramarathon runners. *Free Radic Biol Med* 36:1329–1341 (2004).
- 10 Dessolin J, Schuler M, Quinart A, De Giorgi F, Ghosez L and Ichas F, Selective targeting of synthetic antioxidants to mitochondria: towards a mitochondrial medicine for neurodegenerative diseases? *Eur J Pharmacol* 447:155–161 (2002).
- 11 Gorinstein S, Caspi A, Libman I, Lerner H-Z, Huang D, Leontowich H, et al, Red grapefruit positively influence serum lipids level in patients suffering from coronary atherosclerosis: studies in vitro and in humans. J Agric Food Chem 54:1887-1892 (2006).
- 12 Leontowicz H, Leontowicz M, Drzewiecki J, Jastrzebski Z, Haruenkit R, Poovarodom S, *et al*, Two exotic fruits positively affect rat's plasma composition. *Food Chem* **102**:192–200 (2007).
- 13 Russo GL, Ins and outs of dietary phytochemicals in cancer chemoprevention. *Biochem Pharmacol* 74:533-544 (2007).
- 14 Reddy L, Odhav B and Bhoola KD, Natural products for cancer prevention: a global perspective. *Pharmacol Ther* **99**:1–13 (2003).
- 15 Frankle EN, Lipid oxidation: a review. *Prog Lipid Res* **19**:1–22 (1980).
- 16 Branen AL, Toxicology and biochemistry of butylated hydroxyanisole and butylated hydroxytoluene. J Am Oil Chem Soc 52:59-63 (1975).
- 17 Wanasundara UN and Shahidi F, Canola extracts as an alternative natural antioxidant for canola oil. J Am Oil Chem Soc 71:817–822 (1994).
- 18 Wanasundara PKJPD, Shahidi F and Shukla VKS, Endogenous antioxidants from oilseeds and edible oils. *Food Rev Int* 13:225–292 (1997).

- 19 Gorinstein S, Haruenkit R, Park Y-S, Jung S-T, Zachwieja Z, Jastrzebski Z, et al, Some bioactive compounds and antioxidant potential in fresh and dried Jaffa sweeties: a new kind of citrus fruits. J Sci Food Agric 84:1459–1463 (2004).
- 20 Chen JH and Ho CT, Antioxidant activities of caffeic acid and its related hydroxycinnamic acid compounds. J Agric Food Chem 45:2374–2378 (1997).
- 21 Wattenberg LW, Chemoprevention of carcinogenesis by minor dietary constituents: symposium introduction. *Pharmaceut Biol* 36:6–7 (1998).
- 22 Kathiyar SK and Mukthar H, Tea in chemoprevention of cancer: epidemiologic and experimental studies. Int J Oncol 8:221–238 (1996).
- 23 Yang CS, Chung JY, Yang G, Chhanbra SK and Lee MJ, Tea and tea polyphenols in cancer prevention. *J Nutr* 130:472S-478S (2000).
- 24 Cho JY, Yang SY, Yu SO, Kim BW, Jang HG, Chon SU, *et al*, The actual distributing states of the fresh wild vegetables at five-day traditional markets in Jeonnam district. *Korean J Hortic Sci Technol* 23:396–401 (2005).
- 25 Dar A and Khatoon S, Behavioral and biochemical studies of dichloromethane fraction from the areca catechu nut. *Pharmacol Biochem Behav* 65:1–6 (2000).
- 26 Au TK and Lam TBN, A comparison of HIV-1 integrase inhibition by aqueous and methanol extracts of Chinese medicinal plants. *Life Sci* 68:1687–1694 (2001).
- 27 Lee BC, Jeong YK and Ryu BH, Antioxidative effect of silymarin and silybin purified from *Silybum marianum* on oxidation of human low density lipoprotein by macrophages. *Korean J Appl Microbiol Biotechnol* **25**:286–292 (1997).
- 28 Brand-Williams W, Cuvelier ME and Berset C, Use of free radical method to evaluate antioxidant activity. *Food Sci Technol* 28:25–30 (1995).
- 29 Singleton VL and Rossi JA, A colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. Am J Enol Viticult 16:144–158 (1965).
- 30 Tian Q, Miller EG, Ahmad H, Tang L and Patil BS, Differential inhibition of human cancer cell proliferation by citrus limonoids. *Nutr Cancer* 40:180–184 (2001).
- 31 Velioglu YS, Mazza G, Gao L and Oomah BD, Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *J Agric Food Chem* **46**:4113–4117 (1998).
- 32 Zhou K and Yu L, Total phenolic contents and antioxidant properties of commonly consumed vegetables grown in Colorado. *LWT Food Sci Technol* **39**:1155–1162 (2006).
- 33 Lee SE, Hwang HJ, Ha JS, Ha HS, Jeong HS and Kim JH, Screening of medicinal plant extracts for antioxidant activity. *Life Sci* 73:167–179 (2003).
- 34 Jastrzebski Z, Medina OJ, Moreno LM and Gorinstein S, In vitro studies of polyphenol compounds, total antioxidant capacity and other dietary indices in a mixture of plants (Prolipid). Int J Food Sci Nutr 58:531–541 (2007).
- 35 Turkmen N, Sari F and Velioglu YS, Effects of extraction solvents on concentration and antioxidant activity of black and black mate tea polyphenols determined by ferrous tartrate and Folin–Ciocalteu methods. *Food Chem* **99**:835–841 (2006).
- 36 Jung MJ, Heo S-I and Wang M-H, Free radical scavenging and total phenolic contents from methanolic extracts of Ulmus davidiana. Food Chem 108:482–487 (2008).
- 37 Manosroi J, Dhumtanom P and Manosroi A, Anti-proliferative activity of essential oil extracted from Thai medicinal plants on KB and P338 cell lines. *Cancer Lett* 235:114–120 (2006).